

Research Into the Effect of Proton Pump Inhibitors on Lungs and Leukocytes

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ABSTRACT

Background: Proton pump inhibitors (PPI) are the most commonly used medication in the world. They are prescribed as an effective treatment choice for gastrointestinal system diseases linked to hyperacidity, especially. Additionally, non-indication and unnecessary use are very common. Many publications in recent times have reported significant side effects. However, there are insufficient studies about the mechanism for these side effects.

Methods: Twenty-four Wistar albino rats were used in this study. Rats were divided into 3 groups of control, group-administered H₂ receptor blockers and a group-administered PPI. Medications were administered for 30 days intraperitoneal. After 30 days, rats were euthanized and lung tissue was obtained. Lung was stained for immunohistochemical catalase, superoxide dismutase, Glutathione peroxidase, myeloperoxidase, and toluidine blue and investigated with a light microscope. Transmission electron microscopy (TEM) was used to investigate lung tissues and neutrophil leukocytes. Additionally, lung tissue had biochemical hydrogen peroxide (H₂O₂) levels researched.

Results: H₂O₂ amounts, produced by lysosomes with important duties for neutrophil functions in lung tissues, were found to be statistically significantly reduced in the group-administered PPI. Results from investigations of specimens obtained with immunohistochemical staining observed increases in antioxidant amounts in the PPI group. Investigation with TEM identified more inflammation findings in the lung tissue from the group-administered PPI compared to the control group and the group-administered H₂ receptors.

Conclusion: In conclusion, we identified long-term PPI use disrupts neutrophil leukocyte functions in the lung. All clinicians should be much more careful about PPI use.

Keywords: ARDS, H₂ receptors blockers, immune deficiency, lung, proton pump inhibitors

INTRODUCTION

Proton pump inhibitors (PPI) are among the most commonly used medications in the world. With the initiation of use of the first PPI, omeprazole, at the end of 1980, PPIs were proven to be an effective treatment choice for a variety of diseases linked to acid including gastroesophageal reflux disease, peptic ulcer disease, Helicobacter pylori eradication treatment, dyspepsia, and stress.¹ The use of PPIs rose by 450% toward the end of the 1990s.² Currently, we see many publications related to unnecessary and increasing rates of PPI use. A study in Spain observed a high frequency of PPI use during hospital admission of patients and in the period after discharge.³ Additionally, in the period after discharge, PPI was continued with no indications for nearly 3-6 months. Use to prevent possible harmful effects of a variety of medications on the stomach is excessive. Proton pump inhibitor group medications are known as stomach protectors among

patients. People participating in a survey in research encompassing many countries around the world identified nearly 30% had acid indigestion, heartburn, and reflux complaints.¹ According to the results of the same study, the medications used to treat these people were mainly PPI. People using PPIs for many years are told that these medications only affect the stomach and have no effect on other tissues or organs and that they can use them comfortably. As a result, excessive amounts of use are present.

In the past PPIs were frequently used for patients being treated in intensive care units with frequent multiple drug use. During use by these types of patients, it was predicted to affect patient mortality, so the use of H₂ receptor blockers was recommended in these cases.⁴ It was suggested that deaths caused by the use of PPI in patients during intensive care treatment were associated with bacterial

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growth occurring due to the alkalinity of the stomach pH; bacteria that reach the lungs may lead to infection, and death occurs due to lung infection.^{4,5} However, the H₂ receptor blockers used as alternative treatments also make the stomach pH alkali. Though both medications increase stomach pH, the mechanisms of action are different. The effects of PPIs are shown through acid pH. If the pH of body cells and tissues does not remain within physiologic limits, it is not compatible with life; as a result, if pH does not fall PPIs do not become active and it is proposed they have no effects outside the stomach. It is not fully clear which mechanism forms a variety of side effects of PPIs. In vitro studies have revealed reduced bactericidal activity of neutrophils caused by omeprazole.⁶ Unlike previous explanations related to pneumonia occurring as a result of PPI use and resulting deaths, our thoughts are as follows: the most important mechanism of action in the struggle of leukocytes, especially neutrophil leukocytes, with bacteria involves degradation and digestion of phagocytic bacteria by lysosomes. The pH within lysosomes is very acidic. For degradation and digestion of bacteria within lysosomes, hydrogen peroxide (H₂O₂) should form within lysosomes and the H₂O₂ should later be transformed into hypochlorous acid (HOCl). For the creation of H₂O₂ in lysosomes, H must be taken into the lysosomes. A proton pump is required for the uptake of H by the lysosomes. The proton pumps in lysosomes could be inhibited following long-term treatment with PPI. This suggests that the struggle between neutrophil leukocytes and bacteria would be inefficient.

In light of these findings, in our study, we attempted to research the possible mechanisms of action of PPIs on some tissues and cells outside the stomach and to reveal the cause of side effects assumed to be due to PPI. With this aim, lung tissues from experimental animals were stained with catalase (CAT), myeloperoxidase (MYP), superoxide dismutase (SOD), and glutathione peroxidase (Gpx) stains in an attempt to observe the activities of these enzymes. Additionally, lung tissues were investigated with an electron microscope to assess morphologic changes. These enzymes effective on oxidative stress and antioxidant mechanisms are some of the parameters required to prove our thesis. Superoxide dismutase is a 32 kDa hemodimeric metalloenzyme basically found in plasma, nucleus, and cytosol.⁷ It catalyzes the differentiation of a catalytic copper ion and superoxide radical into dioxygen and H₂O₂. Extracellular SOD is released and synthesized by fibroblasts, glial cells, and endothelial cells. Lung tissue has high extracellular SOD levels. Superoxide dismutase is the only antioxidant that can

inactivate enzymatic free oxygen radicals at the extracellular level. As a result, extracellular SOD undertakes important duties in terms of protecting against diseases like oxidant injury, inflammation, and fibrosis.⁸ The efficacy of SOD is increased when the organism experiences increased oxidant stress. Superoxide dismutase activity is enhanced, especially in clinical situations where reduced effect of antioxidant systems occurs.^{9,10} In our study, in parallel to this information, it was found that SOD activity increased in the lungs.

Glutathione peroxidase is the most effective enzyme in endothelial cells, especially in lungs.¹¹ Free radicals are highly reactive species damaging DNA in proteins, lipids, and carbohydrates causing structural cell injury and apoptosis. Exogenous antioxidant enzymes like CAT, SOD, Gpx, MYP, and thiol groups protect the cells against oxidative stress injury caused by free radicals.¹²

Additionally, we researched whether the H₂O₂ levels found in lung tissues were different between the groups. Clinical research in the recent period has shown significant side effects of this medication group; however, it has still not been fully revealed which mechanism of action is involved in the development of these side effects.

MATERIALS AND METHODS

Experimental Animals and Groups

In this study, 24 Wistar Albino male rats weighing 240-280 g were used. The animals were housed within fixed limits at room temperature (24 ± 2°C) with 55%±15 relative humidity in 12 h/12-h light-dark cycle. Water and standard rat chow were given ad libitum. Care was taken that all procedures related to animals were completed in accordance with national and international regulations related to animal experiments. The study was completed in Kutahya Health Sciences University Experimental Animal Breeding Research and Application Center, Dumlupinar University Advanced Technology Design, Research and Development Center and Eskisehir Osmangazi University, Central Research Laboratory, Application and Research Center. The study was completed in 2 stages. The first study received ethics committee permission from Dumlupinar University Animal Experiments Local Ethics Committee. Later, to more clearly prove the hypothesis, tissue obtained from the previous study was studied with an electron microscope and for this second ethics committee permission was obtained from Kutahya Health Sciences University Animal Experiments Local Ethics Committee.

The first stage of the study was supported by Dumlupinar University—SRP and the second stage was supported by Kutahya Health Sciences University—SRP.

Experimental Animals Were Divided Into 3 Groups

Animals in the First Group (Control Group): Animals in the first group (Control group) were used as controls, were administered a single dose of ip physiological saline with 0.1 cm³ volume every day for 30 days, and were killed at the end of the month with lung tissue removed.

Animals in the Second Group (PPI group): Animals in the second group (PPI group) were administered a single dose of ip PPI (pantoprazole 40 mg vial, Sandoz/Turkey) in a volume of 0.1 cm³, 0.6 mg/kg dose every day for 30 days and were killed at the end of the month with lung tissue removed.

Animals in the Third Group (Ra Group): Animals in the third group (Ra Group) were administered a single dose of ip H₂ receptor blocker (Ranitidin 50 mg/2 ml, Deva/Turkey) in a volume of 0.1 cm³, 4 mg/kg dose every day for 30 days and were killed at the end of the month with lung tissue removed. Lung samples obtained in the study were assessed for histopathologic and biochemical changes.

Histologic Investigation

The removed lung tissues were divided into 2 pieces for histopathologic investigation. Some of the specimens were fixed in 10% neutral formalin solution. Standard histologic techniques were applied. Later they were submerged in paraffin and sectioned to 5 µm thickness. The lung were stained with CAT, SOD, Gpx, and MYP staining and investigated with a light microscope. The section allocated for electron microscope investigation was fixed in 2.5% glutaraldehyde for 24 h at 4°C. Later, secondary fixation was performed in 1% osmium tetroxide at room temperature in a rotator. Tissues were washed 3 times in buffer solution, then passed through an ethyl alcohol series at 4°C twice. Propylene oxide was used for transparency. Tissues were submerged in Araldite the next day. The obtained blocks were sliced with an ultramicrotome and prepared for investigation. Investigation with transmission electron microscopy (TEM) assessed histopathologic changes occurring in lung tissue and especially, changes in neutrophil leukocytes.

All histologic assessments were performed by 2 histologists blinded to the groups.

Immunohistochemical staining in the groups was determined according to intensity (i) as (0): no staining, (1): weak, (2): moderate, (3): strong, and (4) severe staining.

Immune staining histologic scoring system (histologic score: H-score) was calculated with the following equation: H-SCORE: $\sum Pi (i + 1)$. 1. The H-SCOR was separately obtained for lung tissue from the multiplication of staining intensity of stained cells with percentages.¹³

BIOCHEMICAL INVESTIGATION

After lungs were removed from animals, they were homogenized in a mechanical homogenizer in 50 mmol/L phosphate buffer (pH 7.40) for biochemical investigation. Homogenates were centrifuged at 10 000 g for 15 min at 4°C, with supernatants stored at -80°C.

Hydrogen Peroxide Analysis

A hydrogen peroxide kit (ab102500, Abcam) was used according to the manufacturer's instructions and spectrophotometric measurements were performed at 570 nm. H₂O₂ analysis constitutes one of the most important parameters in the study. This method will be assessed to reveal the mechanism of action of the drugs and to prove our hypothesis.

Statistical Analysis

Results are given as mean \pm SEM. Data were assessed with the one-way analysis of variance (ANOVA) test (post hoc Dunnett test) with the SPSS program. The Kruskal-Wallis test (post hoc Dunn's method) was used to compare histopathologic results. $P < .05$ was accepted as significant.

RESULTS

Histologic Investigation Results

Superoxide dismutase: According to results obtained for H-scores for immunohistochemical staining, lungs were observed to have SOD activity increased by a significant degree in the PPI group compared to the control group ($P < .002$). Though there was a significant increase in the Ra group compared to controls, it was not as much as for PPI ($P < .002$) (Figure 1). When this result is compared with immunohistochemical pictures, strong staining was identified in the PPI group, with less staining observed in the Ra and control groups (Figure 2A). According to this

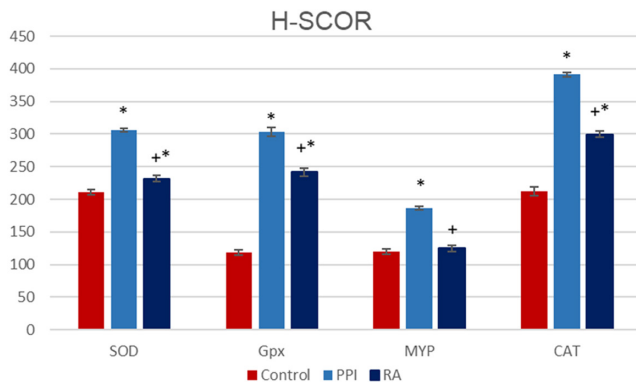


Figure 1. Effects of proton pump inhibitor treatment on histological parameters in rats. Data are as given \pm SD. * $P < .05$ versus control group, + $P < .05$ versus PPI group ANOVA ($n = 8$). SOD, superoxide dismutase; Gpx, glutathione peroxidase; MYP: myeloperoxidase; CAT: catalase; PPI, proton pump inhibitors; Ra: ranitidin.

result, it can be stated that PPIs enhance SOD activity more compared to H2 receptor blockers.

Glutathione Peroxidase: Similar results are encountered for H-score as seen in the SOD group. Glutathione peroxidase activity increased in the PPI group ($P < .002$), with less activity observed in the Ra group ($P < .002$) (Figure 1). When glutathione peroxidase staining is investigated on immunohistochemical pictures, strong immunoreactivity was observed in the PPI and Ra groups, with moderate intensity staining observed in the control group (Figure 2B). PPI were concluded to be more effective on glutathione peroxidase activity in the lung compared to H2 receptor blockers.

Myeloperoxidase: H-scores showed that MYP activity was significantly elevated in the PPI group compared to the control group ($P < .005$). However, the same effect was not observed in the Ra group (Figure 1). When the PPI and Ra groups are compared, MYP values were lower in the Ra group ($P < .005$) (Figure 1). When assessed in terms of myeloperoxidase staining, the PPI group was observed to have moderate severity immunoreactivity, while the Ra and control groups were identified to have less intense staining (Figure 2C).

Catalase: When the H-scores for catalase activity are examined, the means for the groups were observed to be very high. While significant catalase activity was observed in comparing the PPI group with the control group ($P < .002$), the mean in the Ra group was significantly low compared to the PPI group ($P < .002$) but was significantly

high when compared to the control group ($P < .002$) (Figure 1). When the experimental groups were compared in terms of catalase staining, the PPI group had severe immunoreactivity, while the Ra group displayed strong immunoreactivity. The control group was observed to have low-moderate intensity immunoreactivity compared to these 2 groups (Figure 2D).

Lung Toluidin Blue Staining: Light microscope examination revealed pulmonary alveoli with a normal spread in control rat lung samples with toluidine blue staining. Alveolar epithelium had a normal appearance and thin edges. These structures were divided by fine alveolar septa.

According to the microscopic data from the PPI group, increased and abnormal distribution of collagen fibers were observed around blood capillaries. The increase in erythrocyte amounts was notable. Occasionally closed and narrowed alveoli were observed. An alveolar macrophage increase was observed in the alveolar lumen.

According to light microscope data obtained from the group-administered H2 receptor blockers, alveoli and alveoli walls had abnormal and irregular structures. There was very excessive alveolar narrowing. Thickening of alveolar walls was observed (Figure 3).

TEM Investigation Results

In our study, TEM images obtained from the control rat lung samples are presented in Figure 4A-D. According to these data, type 1 and type 2 alveolar epithelial cells and basal lamina in the alveolar wall have a healthy morphology. In general, the alveolar epithelial wall structure showed regularity. Type 1 pneumocytes with a regular oval nucleus in normal sizes and shapes were detected. Intense peripheral chromatin was seen in the nucleus of these pneumocytes. Type 2 pneumocytes with normal peripheral chromatin and regular nucleus structure were also observed. Vacuoles were also found in some cells.

In TEM data obtained from PPI-treated groups (Figure 5A D), the most prominent finding was detected as alveolar epithelial thickening and irregularity. The increase in the amount of erythrocytes is remarkable. The peripheral chromatin density in the cell nucleus increased very much and few erythrocyte debris were observed in the alveolar lumen. In addition, lamellar body increase was observed in type 2 pneumocytes. Nucleus and lamellar body damage and fusion were detected in some type

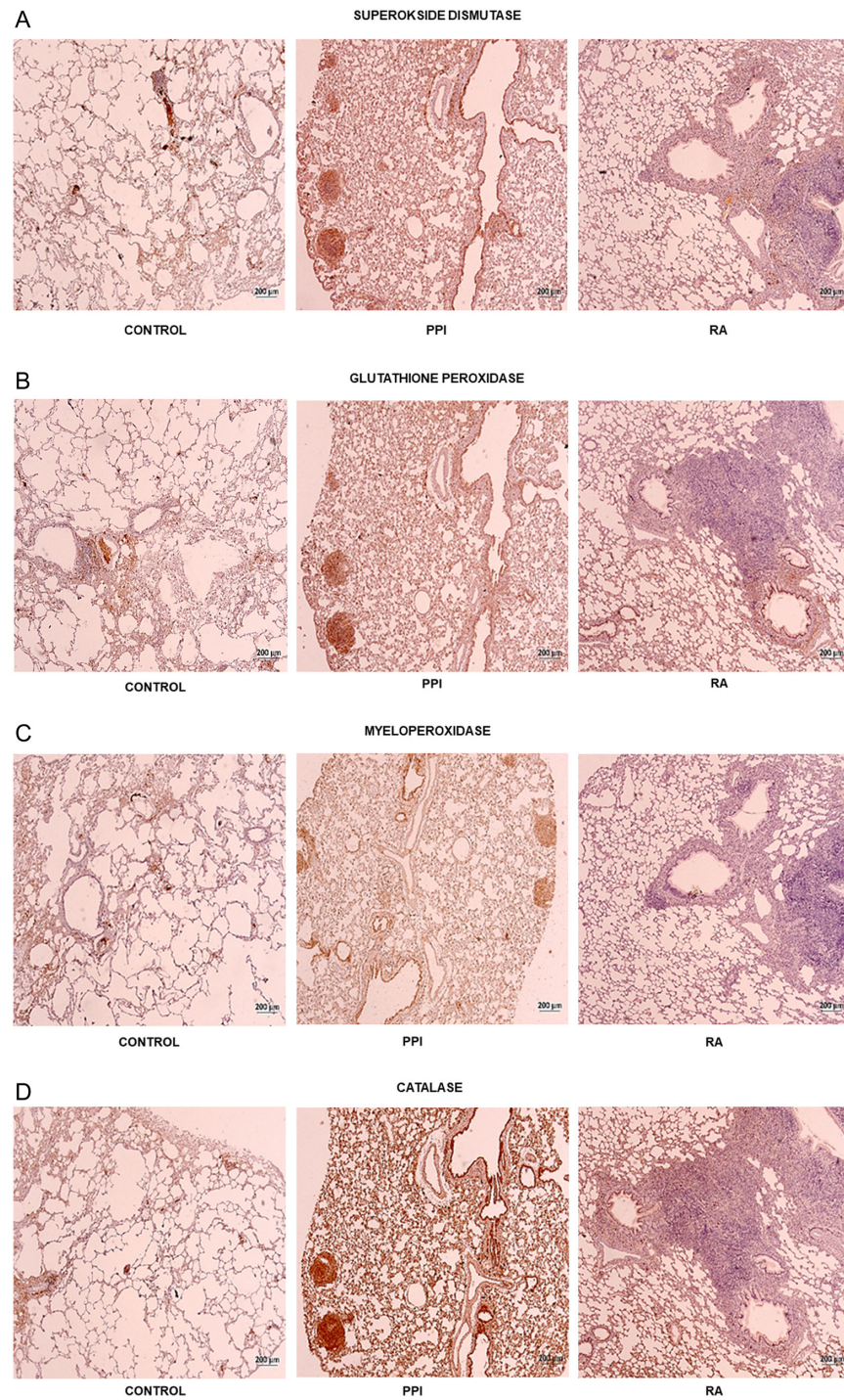


Figure 2. Representative images of immunohistochemical staining. (A) Superoxide dismutase; (B) glutathione peroxidase; (C) myeloperoxidase; (D) catalase.

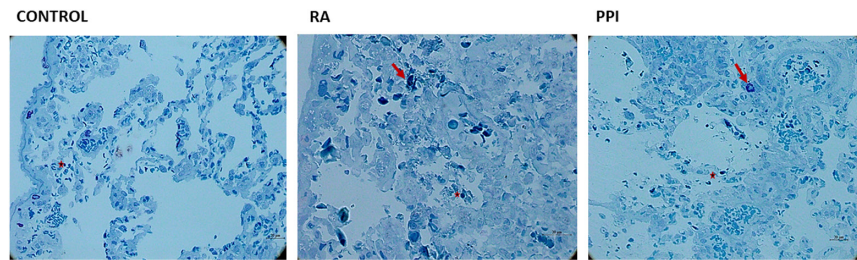


Figure 3. Representative images of lung toluidin blue staining. PPI, proton pump inhibitors; Ra, ranitidin. Arrow: mast cell; Star: leucocyte.

2 cells. Mitochondrial swelling and cristae damage were also observed in some cell cytoplasm. Occasional vacuole formations and electron-dense inclusions were observed. In some nucleus structures, advanced membrane undulations and pyknotic appearance were observed. Cellular debris was observed in some areas of the alveolar lumen (Figure 5A-D).

In TEM micrographs obtained from the H2 receptor blocker treated group alveolar epithelial thickness and integrity loss were observed. In general, the size of the type 2 pneumocytes is larger than the control group, and lamellar bodies were found to be irregular and damaged rather than having a regular oval appearance. Apical microvillus structures were relatively well preserved in some cells. However, there was an irregularity, rupture, and shortening in some areas. In some cells, dense peripheral chromatin

and nucleus undulations were determined. Collagen fibers were evident, mitochondria were relatively healthy, and vacuole formations were observed (Figure 6A-D).

Biochemical Investigation Results

H₂O₂ Analysis: A significant decrease was found with fluorometric measurements in the PPI group. Although this result is not certain, it indicates very low H₂O₂ values in the lung tissue of the group-administered PPI based on average values. While the mean of H₂O₂ value in the control group was 79.33 ± 34.81 , the mean H₂O₂ value in the PPI group was identified as 35.67 ± 17.06 (Figure 7).

DISCUSSION

Lysosomal acid hydrolases, which help the action of lysosomes found in leukocytes in the lungs, are activated at acidic pH. The necessity of acidic pH for lysosomal

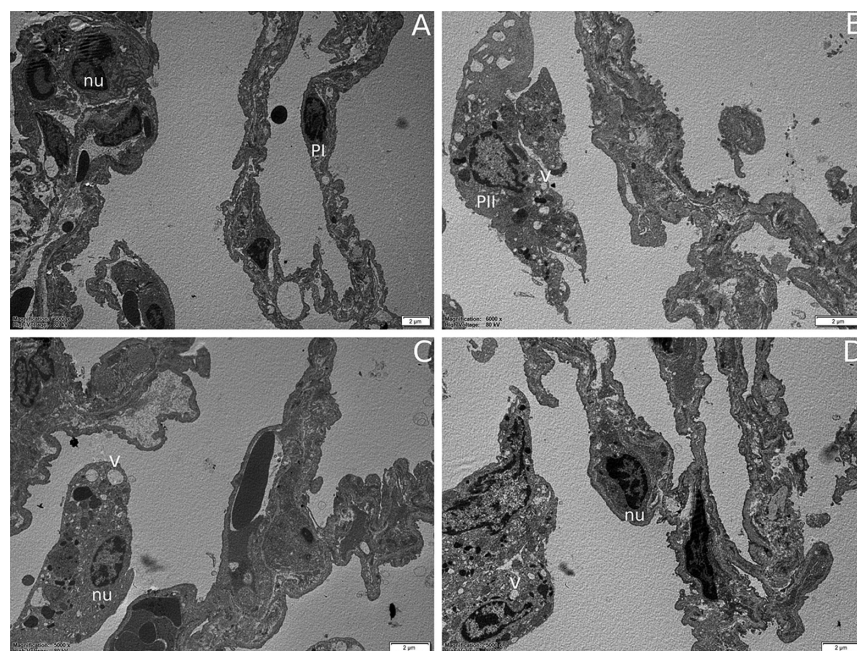


Figure 4. Representative images of lung TEM control group.

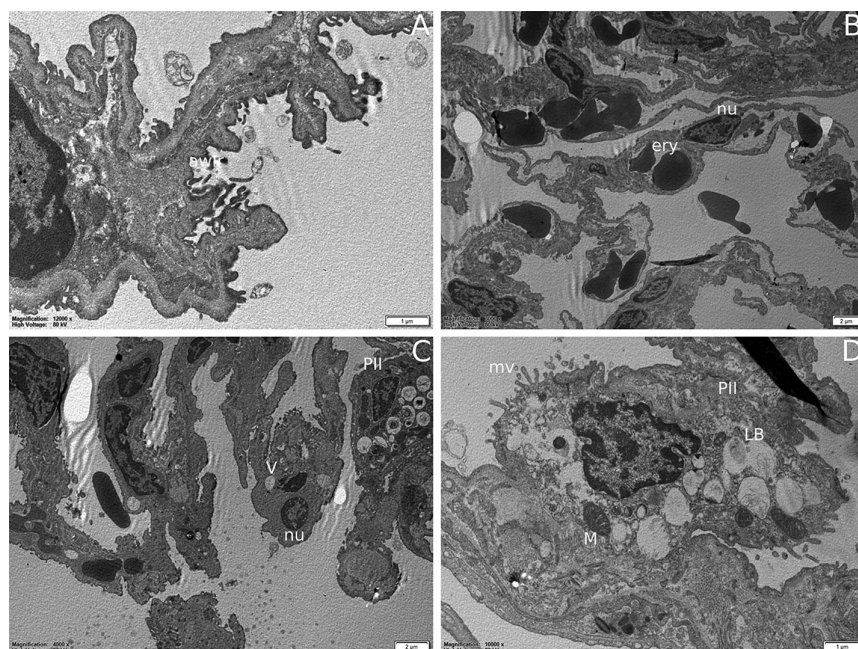


Figure 5. Representative images of lung TEM PPI group.

hydrolases provides double protection against uncontrolled digestion of cytosol content. Even if the lysosomal membrane degrades, the released acid hydrolases will be inactive in the neutral pH of the cytosol. To preserve internal acidic pH, it is necessary for lysosomes to

concentrate active H^+ ions. The active transport of these ions into the cell is assisted by proton pumps in the lysosomal membrane. The most important mechanism of action in the struggle of leukocytes, especially neutrophil leukocytes, with bacteria is the degradation and digestion

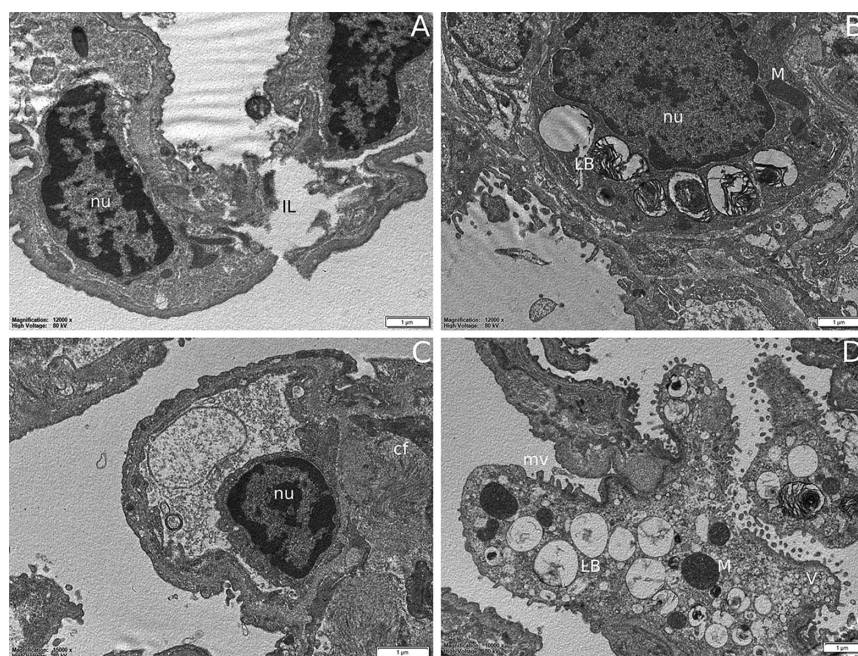


Figure 6. Representative images of lung TEM Ra group.

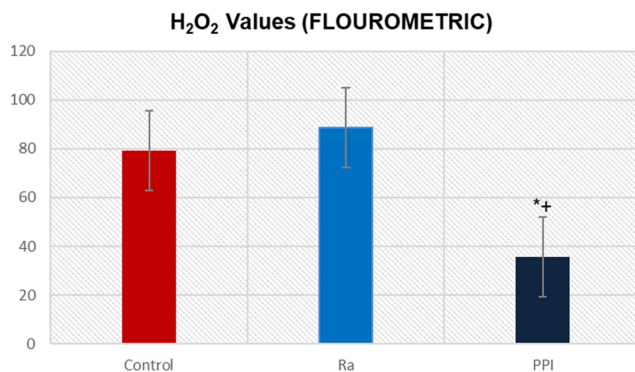


Figure 7. Effects of proton pump inhibitor and H₂ receptor blocker treatment on biochemical parameters in rats. Data are as given \pm SD. * $P < .05$ versus control group, + $P < .05$ versus PPI group ANOVA ($n = 8$). PPI, proton pump inhibitors; Ra: ranitidine.

of phagocytized bacteria with lysosome.¹⁴ For degradation and digestion of bacteria within lysosomes, H₂O₂ must be formed within the lysosome and the H₂O₂ must later transform into HOCl acid.

In conclusion, we observed that the H-indexes of the PPI group were significantly higher than the control group as a result of the examination of SOD, Gpx, MYP, and catalase immunohistochemical staining in lung tissue. The H₂ receptor blocker of ranitidine (Ra) was not as effective as PPI; however, a significant increase was observed in the Ra group compared to the control group. Pictures obtained from groups with histochemical staining and toluidine blue staining showed increased infiltration in lung and histopathologic changes.

Superoxide dismutase is a 32 kDa hemodimeric metalloenzyme basically found in plasma, nuclei, and cytosol.⁷ It catalyzes the separation of a catalytic copper ion and superoxide radical to dioxygen and H₂O₂. Superoxide dismutase activity also increases when the organism experiences increased oxidant stress. Especially in clinical situations with reduced effects of the antioxidant systems, SOD activity increases. In our study, in parallel with this information, SOD activity appeared to be increased in the lung. The reason for the increase in SOD activity is the reduction occurring in oxidative stress, while we think another reason is the catalysis of H₂O₂ formation expected to form in lysosomes. The changes in H₂O₂ amounts support this. The H₂O₂ amount obtained in the PPI group (35.67 ± 17.06) was very low compared to the H₂O₂ amount obtained in the Ra group (88.67 ± 48.28) and this was statistically significant ($P = .022$). In both groups, in spite of histopathologic

injury to the lungs, the statistically significant difference in H₂O₂ amounts found in lung tissue show that PPIs prevent sufficient formation of H₂O₂ in lysosomes found in neutrophils.

In our study, we saw significant degrees of elevation in SOD, CAT, MYP, and Gpx activity in addition to antioxidant enzymes in the control group. A study by Rahimi et al⁹ induced acute respiratory distress syndrome (ARDS) in rats and examined the anti-inflammatory and antioxidant activity of "Portulaca Oleracea" extract and here they identified SOD, CAT, and MYP activity increased after the addition of Portulaca Oleracea extract. The researchers attempted to determine antioxidant enzyme levels with the aid of commercial kits using serum samples. A study by Hackert et al examined the effects of pantoprazole on acute pancreatitis and in conclusion they saw inflammation was suppressed. The researchers concluded that pantoprazole had anti-inflammatory effects and reduced the progression of acute pancreatitis. The researchers examined MPO activity in studies and observed MPO activity increased with pantoprazole.¹⁵ However, a study from 6 years before in 2004 examined the in vivo anti-inflammatory properties of pantoprazole and concluded that it did not have anti-inflammatory features.¹⁶ The antioxidant enzyme activity was increased in the groups given PPI and Ra. The reason for this was not the increase in anti-inflammatory and antioxidant levels due to PPI and ranitidine, it is the activation of the defense mechanism of the body against inflammation and oxidative stress induced by them. The result is an increase in the antioxidant levels in the body.

PPI are the first medications chosen for the treatment of diseases related to acid secretion. Long-term use is observed especially in situations like gastroesophageal reflux. Apart from this, we are against excessive prescriptions in unnecessary situations. In these situations, PPIs affect the lysosome and lysosomes will not be able to fulfill their duty to destroy bacteria and will cause increased lung infection.

In our study results, we observed the thesis related to the proliferation of bacteria linked to increased stomach pH and later pneumonia-causing death, proposed as the reason for deaths occurring as a result of PPI use among patients in intensive care especially, was deficient. Our H₂O₂ results in lung tissue showed that lysosomal function disorder induced in neutrophil leukocytes in lung tissue worsened the progression of the disease.

In conclusion, PPIs do not just affect parietal cells found in the stomach. They may affect proton pumps in other tissues and cells in the organism. As a result, the use and indications for PPI should be reassessed considering more possible side effects than previously determined.

Ethics Committee Approval: This study was approved by the Dumlupinar University Animal Experiments Local Ethics Committee (No: 2018.01.02) and Kutahya Health Sciences University Animal Experiments Local Ethics Committee. (No: 2019.05.02).

Informed Consent: N/A.

Peer-review: Externally peer-reviewed.

Author Contributions: Concept – O.O., F.Y.O.; Design – O.O., F.Y.O., Y.T.; Supervision – I.D., O.O.; Resource – F.Y.O., A.K., S.S., Y.T.; Materials – O.O., I.D., F.Y.O.; Data Collection and/or Processing – S.S., Y.T., F.Y.O., O.O., I.D.; Analysis and/or Interpretation – O.O., F.Y.O., A.K., Y.T.; Literature Search – O.O., A.K., S.S., F.Y.O.; Writing – O.O., F.Y.O.; Critical – I.D., A.K., Y.T.; Reviews – I.D., O.O., Y.T., F.Y.O., S.S.

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Conflict of Interest: The authors declare that they have no conflict of interest.

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REFERENCES

1. Luo H, Fan Q, Xiao S, Chen K. Changes in proton pump inhibitor prescribing trend over the past decade and pharmacists' effect on prescribing practice at a tertiary hospital. *BMC Health Serv Res.* 2018;18(1):537. [\[CrossRef\]](#)
2. Guda NM, Noonan M, Kreiner MJ, Partington S, Vakil N. Use of intravenous proton pump inhibitors in community practice: an explanation for the shortage? *Am J Gastroenterol.* 2004;99(7):1233-1237. [\[CrossRef\]](#)
3. Ramirez E, Lei SH, Borobia AM, et al. Overuse of PPIs in patients at admission, during treatment, and at discharge in a tertiary Spanish Hospital. *Curr Clin Pharmacol.* 2010;5(4):288-297. [\[CrossRef\]](#)
4. Hamai K, Iwamoto H, Ohshimo S, et al. Use of proton pump inhibitors is associated with increased mortality due to nosocomial pneumonia in bedridden patients receiving tube feeding. *Geriatr Gerontol Int.* 2018;18(8):1215-1218. [\[CrossRef\]](#)
5. Garvey BM, McCambley JA, Tuxen DV. Effects of gastric alkalization on bacterial colonization in critically ill patients. *Crit Care Med.* 1989;17(3):211-216. [\[CrossRef\]](#)
6. Zedtwitz-Liebenstein K, Wenisch C, Patruta S, et al. Omeprazole treatment diminishes intra- and extracellular neutrophil reactive oxygen production and bactericidal activity. *Crit Care Med.* 2002;30(5):1118-1122. [\[CrossRef\]](#)
7. Chen L, Watson C, Morsch M, et al. Improving the delivery of SOD1 antisense oligonucleotides to motor neurons using calcium phosphate-lipid nanoparticles. *Front Neurosci.* 2017;30(11):476. [\[CrossRef\]](#) eCollection.
8. Gao F, Kinnula VL, Myllärniemi M, Oury TD. Extracellular superoxide dismutase in pulmonary fibrosis. *Antioxid Redox Signal.* 2008;10(2):343-354. [\[CrossRef\]](#)
9. Baradaran Rahimi VB, Rakhshandeh H, Raucci F, et al. Anti-inflammatory and anti-oxidant activity of *Portulaca oleracea* Extract on LPS-induced rat lung injury. *Molecules.* 2019;24(1):139. [\[CrossRef\]](#)
10. Limón-Pacheco J, Gonsébat ME. The role of antioxidants and antioxidant-related enzymes in protective responses to environmentally induced oxidative stress. *Mutat Res.* 2009;674(1-2):137-147. [\[CrossRef\]](#)
11. Cheeseman KH, Slater TF. An introduction to free radical biochemistry. *Br Med Bull.* 1993;49(3):481-493. [\[CrossRef\]](#)
12. Sheng Y, Abreu IA, Cabelli DE, et al. Superoxide dismutases and superoxide reductases. *Chem Rev.* 2014;114(7):3854-3918. [\[CrossRef\]](#) Epub 2014 Apr 1.
13. McCarty KS Jr, Miller LS, Cox EB, Konrath J, McCarty KS Sr. Estrogen receptor analyses. Correlation of biochemical and immunohistochemical methods using monoclonal antireceptor antibodies. *Arch Pathol Lab Med.* 1985;109(8):716-721.
14. Cooper GM. Lysosomes. In: 2nd ed Cooper G. M, ed. *The Cell* Sunderland (MA) Sinauer Associates; Sunderland; 2000.
15. Hackert T, Tudor S, Felix K, et al. Effects of pantoprazole in experimental acute pancreatitis. *Life Sci.* 2010;87(17-18):551-557. [\[CrossRef\]](#)
16. Becker TL, Maróstica M, Ribeiro ML, et al. Pantoprazole treatment does not invoke anti-inflammatory properties in vivo. *Int Immunopharmacol.* 2004;4(8):1051-1057. [\[CrossRef\]](#)